# Diterpenoid Alkaloids from Aconitum leucostomum

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Two new norditerpenoid alkaloids, leucostines A (1) and B (2), and a new diterpenoid alkaloid, 11-acetyllepenine (6), have been isolated from the root of *Aconitum leucostomum*. Their structures were elucidated on the basis of spectral data and chemical evidence. Four known alkaloids, delsoline (3), delcosine (4), lepenine (7), and songorine (8), were also isolated and identified from this plant for the first time.

Aconitum leucostomum Vorosch. (Ranunculaceae) is a perennial herb distributed in the Gansu and Xinjing provinces of China. It has been used as a folk medicine to treat traumatic injury. The isolation of several alkaloids from this plant has been reported.<sup>1–6</sup> This paper describes the isolation and structure elucidation of three new diterpenoid alkaloids, named leucostines A (1) and B (2) and 11-acetyllepenine (6), as well as the isolation of four known alkaloids, delsoline (3), delcosine (4), lepenine (7), and songorine (8), from this plant. The latter were isolated from this plant for the first time.

Leucostine A (1), C<sub>27</sub>H<sub>43</sub>NO<sub>8</sub>, is an amorphous powder. The IR spectrum indicated the presence of hydroxyl (3542 cm<sup>-1</sup>) and ester (1735 cm<sup>-1</sup>) groups. The <sup>1</sup>H NMR revealed the presence of four methoxy groups (3.43, 3.38, 3.31, and 3.28 ppm, each 3H, s), an acetyl group (2.21, 3H, s), and an ethylamine group (1.06, 3H, t, J = 7.1 Hz). In the <sup>13</sup>C NMR, 27 signals appeared, corresponding to 27 carbon atoms in the molecule. These spectral data suggested that the chemical formulation of this compound was a norditerpenoid alkaloid, C<sub>19</sub>H<sub>21</sub>(OH)<sub>2</sub>(OCH<sub>3</sub>)<sub>4</sub>(OAc)(NCH<sub>2</sub>CH<sub>3</sub>). Most alkaloids of this class have a hydroxyl or a methoxyl group on C-1, C-8, C-14, and C-16. As no angular methyl group was observed in the <sup>1</sup>H-NMR or <sup>13</sup>C-NMR spectrum, the C-18 most likely bears an oxygen function. The secondary carbon at  $\delta$  78.57 ppm indicates that the C-18 methylene carbon bears a methoxyl group. In the case of a hydroxyl group on C-18, the signal normally appears at about 66.5-68.5 ppm.<sup>7,8</sup> The tertiary carbons at  $\delta$  83.95 and 81.06 ppm indicate that both the C-1 and C-16 bear a methoxyl group. The signal at 3.74 ppm (1H, t, J = 5.3 Hz, C-14- $\beta$ -H) suggests that C-13 and C-9 did not bear hydroxyl groups. It also appears that C-10 did not bear an oxygen function. This conclusion is supported by the quaternary carbon signal at  $\delta$  48.39 ppm assigned to C-11. In the case in which an oxygen group was located on the C-10, the C-11 signal appeared near  $\delta$  55–56 ppm<sup>9</sup> and the C-14- $\beta$ -H signal was shifted more downfield. 10 Quaternary carbon signals at  $\delta$  88.87 and 76.33 ppm showed that C-7 and C-8 were connected by a hydroxyl group. The <sup>1</sup>H-NMR signal at  $\delta$  5.34 ppm did not disappear in the D<sub>2</sub>O exchange experiment, suggesting that C-6 was connected with a  $\beta\text{-OAc.}^{11-13}$  This conclusion was supported by the  $^{13}\text{C-NMR}$  signal at  $\delta$  81.06 ppm assigned to C-6. The proton signal at  $\delta$  3.74 ppm (1H, t, J=5.3 Hz) for C-14- $\beta$ -H and the carbon signal at  $\delta$  84.31 ppm for C-14 indicated that C-14 bears an  $\alpha\text{-OMe.}$  Comparison of the  $^{13}\text{C-NMR}$  data of 1 with those of the known alkaloid delphatine (5) $^{17}$  showed that the chemical shifts of carbons were very similar except for the differences due to different substituents at C-6. The structure of leucostine A was thus assigned to be 1.

Leucostine B (2), crystallized from n-hexane-Me<sub>2</sub>CO as colorless needles, mp 263-264 °C, has the molecular formula C<sub>24</sub>H<sub>39</sub>NO<sub>8</sub> (HRMS). Its IR spectrum indicated the presence of hydroxyl groups (3477–3374 cm<sup>-1</sup>). The presence of an ethylamine group was inferred by the proton resonances at  $\delta$  1.12 ppm (3H, t, J = 7.0 Hz) in the <sup>1</sup>H-NMR spectrum. The <sup>1</sup>H NMR also indicated the presence of three methoxy groups (3.40, 3.37, and 3.32, s, each 3H). The HRMS and NMR analyses suggested that the chemical formula of leucostine B (2) was C<sub>19</sub>H<sub>20</sub>(OCH<sub>3</sub>)<sub>3</sub>(OH)<sub>5</sub>(NCH<sub>2</sub>CH<sub>3</sub>), corresponding to that of a norditerpenoid alkaloid. The secondary carbon signal at  $\delta$  77.21 ppm indicated that the C-18 methylene carbon bore a methoxyl group. Quaternary carbon signals at  $\delta$  37.78 and 48.00 ppm was assigned to C-4 and C-11, respectively. The other two quaternary carbon signals at  $\delta$  77.34 and 87.90 ppm in the  $^{13}C-$ NMR spectrum could be assigned to any of the carbons bearing hydroxyls, such as C-7, C-8, C-9, C-10, and C-13. The C-10 position can be excluded, as the C-11 signal appears at  $\delta$  48.00 ppm rather than near  $\delta$  55–56 ppm, as would be expected. The C-14- $\beta$ -H appeared as a quartet in the <sup>1</sup>H NMR, indicating that C-9 or C-13 did not bear a hydroxyl. Leucostine B (2) should, therefore, bear hydroxyls at C-7 and C-8, and these carbon signals are observed at  $\delta$  87.90 and 77.34 ppm, respectively. The tertiary signals at  $\delta$  89.93 and 78.91 ppm were, respectively, assigned to C-6 and C-16 bearing a  $\beta$ -OMe. The signal at  $\delta$  73.57 ppm indicated that an  $\alpha$ -OH was located at C-1. The proton signal at 4.51 ppm (1H, q, J= 4.7 Hz) and the carbon signal at  $\delta$  75.69 ppm showed that an  $\alpha$ -OH was located at C-14. The above spectral evidence leads to the partial structure of leucostine B, which is the same as delcosine (4). The remaining hydroxyl group can be at C-2, C-3, C-12, or C-15,

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because the tertiary signal at  $\delta$  71.90 ppm can only be assigned to one of these carbons bearing a hydroxyl group. Comparison of the <sup>13</sup>C-NMR data of **2** with those of the known alkaloid delcosine (4) showed that the chemical shifts of carbons C-1, C-2, C-3, and C-4 of 2 were very similar to those of delcosine (4). Thus, location of a hydroxy group at C-2 or C-3 was ruled out. A hydroxy group at C-15 is unlikely because the chemical shifts of C-8 and C-15 are very close to those of delcosine (4). The C-16 ( $\delta$  78.91 ppm) of **2** compared with that of delcosine (4) ( $\delta$  81.97 ppm) was shifted upfield about 3 ppm. This is a shift opposite of that expected for a hydroxy group at C-15. The remaining hydroxyl group was thus definitely assigned to C-12. The C-13 ( $\delta$  55.59 ppm) and C-10 ( $\delta$  49.42 ppm) signals compared with those of delcosine (4) were shifted downfield about 10 ppm as a result of the deshielding of the C-12 hydroxyl group. The chemical shift of C-16 ( $\delta$  78.91 ppm) moved upfield about 3 ppm from that in delcosine ( $\delta$  81.97 ppm) as a result of the  $\gamma$ -gauche effect of the C-12 hydroxyl group, suggesting that the C-12-OH is  $\alpha$ -oriented. If the C-12-OH were  $\beta$ -oriented, it would have a  $\gamma$ -gauche relation with C-14. The chemical shift of the C-14 ( $\delta$  75.51 ppm) of **2** is very close to that of delcosine (4) ( $\delta$  75.67 ppm), supporting the C-12-OH α-orientation. Structure 2 was thus assigned to leucostine B.

11-Acetyllepenine (6) was obtained as colorless prisms, mp 130-131 °C. Its formula was inferred as C24H35-NO<sub>4</sub> by the <sup>13</sup>C NMR and EIMS. The IR spectrum showed the presence of hydroxyl (3387 cm<sup>-1</sup>) and ester (1735 cm<sup>-1</sup>) functions. Signals at  $\delta$  5.23 and 4.97 ppm (each 1H, d, J = 2.2 Hz, H-17), 1.05 (3H, t, J = 7.5 Hz, -NCH<sub>2</sub>CH<sub>3</sub>), and 0.70 ppm (3H, s, C-4-CH<sub>3</sub>) in the <sup>1</sup>H NMR indicated that compound 6 was a diterpenoid alkaloid. Analysis of the <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of compound 6 and lepenine (7) showed that both compounds possess the atisine skeleton and differ in the substituent at C-11. The quaternary carbon signals at  $\delta$  33.67, 43.53, and 50.99 ppm were assigned to C-4, C-8, and C-10, respectively. The tertiary carbon signals at  $\delta$  70.21 and 77.61 ppm were assigned to the C-1 and C-15 bearing a hydroxyl, respectively. The other tertiary carbon signal at  $\delta$  76.17 ppm was assigned to C-11 bearing an acetyl group. Comparison of the <sup>1</sup>H NMR of 6 with that of lepenine (7) showed that the <sup>1</sup>H-NMR signal of **6** at  $\delta$  5.52 ppm (1H, d, J = 9.1 Hz, C-11- $\alpha$ -H) was shifted upfield to 4.44 ppm (1H, d, J = 9.1 Hz, C-11α-H) in the <sup>1</sup>H NMR of lepenine (7), further evidence that the acetyl was located at C-11. Thus, structure **6** was identified as 11-acetyllepenine. This conclusion was confirmed by hydrolysis of 6 with 5% KOH-aqueous MeOH to give lepenine (7).15

The other four known alkaloids isolated along with these new alkaloids were identified as delsoline (3),  $^{16,17}$  delcosine (4),  $^{17,18}$  lepenine (7),  $^{15}$  and songorine (8) on the basis of their physical constants and spectral data.

### **Experimental Section**

**General Experimental Procedures.** Melting points were obtained on a Kofler apparatus and are uncorrected. IR spectral data were measured on a FT-5DX instrument with KBr disks. EIMS data were obtained on a VGZAB-HS mass spectrometer. <sup>1</sup>H NMR and <sup>13</sup>C NMR were recorded on a Bruker AM-400 instrument with TMS as an internal standard and CDCl<sub>3</sub> as solvent.

- 1: R<sub>1</sub>=R<sub>3</sub>=CH<sub>3</sub>,R<sub>2</sub>=Ac, R<sub>4</sub>=H
- 2: R<sub>1</sub>=R<sub>3</sub>=H, R<sub>2</sub>=CH<sub>3</sub>, R<sub>4</sub>=OH
- 3: R<sub>1</sub>=R<sub>4</sub>=H, R<sub>2</sub>=R<sub>3</sub>=CH<sub>3</sub>
- 4: R<sub>1</sub>=R<sub>3</sub>=R<sub>4</sub>=H, R<sub>2</sub>=CH<sub>3</sub>
- 5: R<sub>1</sub>=R<sub>2</sub>=R<sub>3</sub>=CH<sub>3</sub>, R<sub>4</sub>=H

**Plant Material.** Aconitum leucostomum was collected from Xinjiang Province, China, in September 1990. It was identified by Prof. Ru-Neng Zhao, Department of Pharmacy, Lanzhou Medical College. A voucher specimen has been deposited in the Department of Biology, Lanzhou University.

**Extraction and Isolation.** The powdered herb (2.5 kg) was extracted with 95% EtOH three times (each time for 4 days) at room temperature. After removal of EtOH under reduced pressure, 85 g of syrup remained. This was dissolved in 2% H<sub>2</sub>SO<sub>4</sub> solution. The acidic solution, after extraction with CH<sub>2</sub>Cl<sub>2</sub>, was made alkaline with concentrated NH<sub>4</sub>OH, adjusted to pH 11, and then extracted with CHCl<sub>3</sub> to give 12.5 g of crude alkaloid. The crude alkaloid (12.5 g) was chromatographed on a column of alumina (250 g) eluting with CHCl<sub>3</sub> and CHCl<sub>3</sub>-CH<sub>3</sub>OH (40:1, 30:1, 20:1, 10:1, and 5:1) to afford 21 fractions. Fractions 1 and 2 were combined and chromatographed on an alumina column eluting with petrol-Et<sub>2</sub>O to give leucostine A (1) and 11acetyllepenine (6). Fractions 4-6 were combined and crystallized from hexane-Me<sub>2</sub>CO to give songorine (8). Fraction 7 was chromatographed on an alumina column eluting with petroleum-Me<sub>2</sub>CO to afford delsoline (3). Fractions 8–13 were combined and chromatographed on an alumina column eluting with petroleum ether-Me<sub>2</sub>CO to afford delcosine (4) and lepenine (7). Fractions 16-20 were combined and chromatographed on an alumina column with CHCl<sub>3</sub>-MeOH (6:1) as eluent to yield leucostine B (2).

Conversion of 11-Acetyllepenine (6) to Lepenine (7). A mixture of 30 mg of 6 in 5 mL of KOH-MeOH solution was kept for 3 days at room temperature. After removal of MeOH under reduced pressure, a small amount of  $H_2O$  was added, and the mixture was extracted with  $CHCl_3$ . The extract was crystallized from hexane-Me<sub>2</sub>CO to yield 20 mg of needle crystals whose physical constants and spectral data were in good agreement with those of lepenine.

**Leucostine A (1):** C<sub>27</sub>H<sub>43</sub>NO<sub>8</sub> (HRMS, M - OMe, found 478.2738, requires 478.2804), was isolated as an amorphous powder; UV (MeOH)  $\lambda$  max (nm) 248.5; IR  $\nu$  max (KBr) cm<sup>-1</sup> 3542 (-OH), 1735 (-COMe);  $^1$ H NMR (CDCl<sub>3</sub> 400 MHz)  $\delta$  (ppm) 5.34 (1H, s, C-6-α-H), 3.74 (1H, t, J = 5.3 Hz, C-14- $\beta$ -H), 3.43, 3.38, 3.31, and 3.28 (each 3H, s, -OMe), 2.21 (3H, s, -COMe), 1.06 (3H, t,

**Table 1.** <sup>13</sup>C-NMR Data of Compounds **1**, **2**, and **6** (CDCl<sub>3</sub>)<sup>a</sup>

carbons	1	2	6
1	83.95	73.57	70.21
2	25.96	28.64	30.95
3	31.79	29.49	38.52
4	38.56	37.78	33.67
5	43.24	$45.80^{b}$	49.02
6	81.06	89.93	23.51
7	88.87	87.90	43.26
8	76.33	77.34	43.53
9	51.53	$45.94^{b}$	51.96
10	37.27	49.42	50.99
11	48.39	48.00	76.17
12	28.73	71.90	42.02
13	45.60	55.60	23.97
14	84.31	75.51	37.15
15	37.93	34.76	77.61
16	82.12	78.91	153.78
17	66.08	66.96	109.57
18	78.57	77.21	25.91
19	52.67	56.17	56.59
20			67.59
$NCH_2CH_3$	51.17	50.59	50.73
	14.18	13.73	13.50
$COCH_3$	172.47		171.09
	21.54		21.51
$C_1$ -OCH $_3$	55.64		
$C_6$ -OCH $_3$		57.43	
$C_{14}$ -OCH <sub>3</sub>	57.67		
$C_{16}$ -OCH <sub>3</sub>	56.25	56.40	
$C_{18}$ -OCH <sub>3</sub>	59.41	59.08	

 $<sup>^</sup>a$  Assignment are partly based on the DEPT techniques.  $^b$  Assignments in the same column may be interchanged.

J = 7.1 Hz,  $-NCH_2CH_3$ ); EIMS m/z 509 [M]<sup>+</sup> (5), 478 [M - OMe]<sup>+</sup> (100), 450 [M - OAc]<sup>+</sup> (30); <sup>13</sup>C NMR, see Table 1.

**Leucostine B (2):** C<sub>24</sub>H<sub>39</sub>NO<sub>8</sub> (HRMS, found 469.2793, requires 469.2776), obtained as colorless needles (MeOH): mp 263–264 °C; IR  $\nu$  max (KBr) cm<sup>-1</sup> 3374–3477 (–OH); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  (ppm) 4.51 (1H, q, J = 4.7 Hz, C-14- $\beta$ -H), 4.11 (1H, d, J = 2.4 Hz, C-6- $\alpha$ -H), 3.40, 3.37, 3.32 (each 3H, s, –OMe), 1.12 (3H, t, J = 7.0 Hz, –NCH<sub>2</sub> $CH_3$ ); EIMS m/z 469 [M]<sup>+</sup> (15), 4.54 [M – CH<sub>3</sub>]<sup>+</sup> (100), 438 [M – OMe]<sup>+</sup> (70); <sup>13</sup>C NMR, see Table 1.

**Delcosine (4)** was obtained as colorless prisms (Me<sub>2</sub>CO-n-hexane): mp 204–205 °C. IR  $\nu$  max (KBr) cm<sup>-1</sup> 3585, 3473, and 3325 (–OH). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  (ppm) 7.33 (1H, d, J = 8.1 Hz, C-1- $\alpha$ -OH), 4.12 (1H, d, J = 4.8 Hz, C-14- $\beta$ -H), 3.67 (1H, br d, J = 7.7 Hz, C-6- $\alpha$ -H), 3.41, 3.40, and 3.33 (each 3H, s, –OMe), 1.10 (3H, t, J = 7.2 Hz, –NCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz)  $\delta$  72.62 (C-1), 27.41 (C-2), 29.32 (C-3), 37.54 (C-4), 43.91 (C-5), 89.99 (C-6), 87.79 (C-7), 78.03 (C-8), 45.25 (C-9), 39.36 (C-10), 48.79 (C-11), 29.32 (C-12), 45.15 (C-13), 75.69 (C-14), 34.39 (C-15), 81.97 (C-16), 66.33 (C-17), 77.27 (C-18), 57.07 (C-19), 50.39, 13.54 (–NEt), 57.32 (C-6-OMe), 56.34 (C-16-OMe), and 59.06 (C-18-OMe); EIMS m/z 453 [M]<sup>+</sup> (13), 438 [M – Me]<sup>+</sup> (100), 422 [M – OMe]<sup>+</sup> (55).

**11-Acetyllepenine (6),** C<sub>24</sub>H<sub>35</sub>NO<sub>4</sub>, was obtained as colorless prisms (Me<sub>2</sub>CO–n-hexane): mp 130–131 °C; UV  $\lambda$  max (nm) 246.5; IR  $\nu$  max (KBr) cm<sup>-1</sup> 3387 (–OH), 1735 (–COMe), 1652 (–C=CH<sub>2</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  (ppm) 5.52 (1H, d, C-11- $\alpha$ -H), 5.23, and 4.97 (each 1H, d, J = 2.2 Hz, C-17-H), 4.32 (1H, d, J = 2.2 Hz, C-15- $\alpha$ -H), 3.85 (1H, dd, J = 5.0 Hz, C-1- $\beta$ -H), 2.08 (3H, s, –OCOMe), 1.05 (3H, t, J = 7.5 Hz, –NCH<sub>2</sub>CH<sub>3</sub>), 0.70 (3H, s, C-4-Me); EIMS m/z 401 [M]<sup>+</sup> (40), 384 [M-OH]<sup>+</sup> (10), 342 [M-OAc]<sup>+</sup> (87); <sup>13</sup>C NMR, see Table 1.

**Lepenine (7)** was obtained as colorless prisms (Me<sub>2</sub>CO-n-hexane): mp 120-122 °C, IR  $\nu$  max (KBr) cm<sup>-1</sup> 3383 (-OH), 1655 (-C=CH<sub>2</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  (ppm) 5.25 and 5.02 (each 1H, d, J = 2.2 Hz, C-17-H), 4.44 (1H, d, J = 9.1 Hz, C-11- $\alpha$ -H), 4.27 (1H, d, J = 2.2 Hz, C-15- $\alpha$ -H), 4.16 (1H, dd, J = 6.2 Hz, C-1- $\beta$ -H), 1.06 (3H, t, J = 7.1 Hz, -NCH<sub>2</sub>CH<sub>3</sub>), 0.70 (3H, s, C-4-Me); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz)  $\delta$  (ppm) 70.64 (C-1), 31.10 (C-2), 38.65 (C-3), 33.71 (C-4), 53.82 (C-5), 23.06 (C-6), 46.85 (C-7), 43.68 (C-8), 52.31 (C-9), 50.98 (C-10), 72.95 (C-11), 42.29 (C-12), 34.61 (C-13), 27.37 (C-14), 77.87 (C-15), 154.37 (C-16), 109.40 (C-17), 25.96 (C-18), 57.02 (C-19), 67.72 (C-20), and 50.79, 13.54 (-NEt); EIMS m/z 359 [M]<sup>+</sup> (50), 342 [M - OH]<sup>+</sup> (81), and 43 (100).

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